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(54) Title: THE USE OF A PROTEIN TYROSINE INHIBITOR SUCH AS GENISTEIN IN THE TREATMENT OF DIABETIC RETINOPATHY OR OCULAR INFLAMMATION		
(57) Abstract The present invention is directed to a method for the prophylactic and therapeutic treatment of diabetic retinopathy and a method for the prophylactic and therapeutic treatment of ocular inflammation. The methods involve the administration of an inhibitor of the protein tyrosine kinase pathway to an animal, such as a mammal, in particular a human, in an amount sufficient to treat diabetic retinopathy or ocular inflammation, respectively, prophylactically or therapeutically. The inhibitor of the protein tyrosine kinase pathway is preferably genistein or an analogue or prodrug thereof or a pharmaceutically acceptable salt of any of the foregoing.		

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THE USE OF A PROTEIN TYROSINE INHIBITOR SUCH AS GENISTEIN IN THE TREATMENT OF DIABETIC RETINOPATHY OR OCULAR INFLAMMATION

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TECHNICAL FIELD OF THE INVENTION

The present invention relates to a method for the prophylactic and therapeutic treatment of diabetic retinopathy and a method for the prophylactic and therapeutic treatment of ocular inflammation.

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BACKGROUND OF THE INVENTION

More than 14 million people in the United States have diabetes. All people with diabetes are at risk of retinal complications. However, people with type I, i.e., insulin-dependent, diabetes, face a greater risk of severe vision loss than people with type II, i.e., non-insulin dependent, diabetes.

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Initially, the high blood glucose level in diabetic people causes an increase in growth factors in their eyes. This condition is known as the "pre-diabetic retinopathy stage" and can lead to retinopathy, if not prophylactically treated.

Retinopathy will affect the majority of diabetic people to some extent during their lifetimes (Anonymous, *MMWR* 42(10): 191-195 (1993)). It is the leading cause of blindness in Americans of age 20 to 74 today and is expected to impair vision in approximately one-third of diabetic people in the United States. Each year in the United States, as many as 40,000 new cases of blindness occur among diabetic people (CDC, unpublished data, 1993). Diabetic people are 25 times more likely than the general population to become blind due to retinopathy.

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Diabetic retinopathy has two stages -- a nonproliferative stage, which typically occurs first, and a proliferative stage. The nonproliferative stage, which is also referred to as "background diabetic retinopathy," is characterized by thickening of the basement membrane, loss of retinal pericytes, microvascular abnormalities, intraretinal microaneurysms, retinal hemorrhages (also known as "dot blot" hemorrhages), retinal edema, in particular diabetic macular edema, capillary closure associated with retinal ischemia or poor retinal perfusion (i.e., poor vessel

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development) and soft and hard exudates. The proliferative stage, which affects an estimated 700,000 Americans (Chen et al., *J. Miss. State Med. Assoc.* 36(7): 201-208 (1995)), is characterized by neovascularization and fibrovascular growth (i.e., scarring involving glial and fibrous elements) from the retina or optic nerve over the inner
5 surface of the retina or disc or into the vitreous cavity.

The proliferative stage can lead to rubeotic or neovascular glaucoma. Macular edema can occur in either stage and it, along with complications from retinal neovascularization, are the two major retinal problems that cause the diabetes-related vision loss.

10 While the pathological stages of diabetic retinopathy are well-described, the molecular events underlying diabetic retinopathy are poorly understood. This is due, in part, to the fact that the disease progresses over ten to thirty years, depending on a given individual.

Tight control of glycemia and hypertension and ophthalmic screening of
15 diabetics appears beneficial in preventing the disease. Current treatment consists of regular observation by an ophthalmologist, laser photocoagulation and vitrectomy.

Ocular inflammation, e.g., macular edema, can occur as a result of disease, bacterial or viral infection, ocular surgery, which includes cataract surgery, retinal surgery, refractive surgery, and corneal surgery, e.g., corneal transplantation, and the
20 like. Ocular inflammation can also occur as a result of corneal transplant rejection.

Ocular inflammation is currently treated by corticosteroids. The disadvantages of such treatment are cataracts, increased intraocular pressure, corneal melting, systemic complications, high blood pressure, aseptic necrosis of the femoral head, cardiovascular diseases, facial hair and many others. Nonsteroidal anti-inflammatory
25 compounds are also administered. The disadvantages of such compounds include certain blood diseases, peptic ulcers, gastric distress, and platelet dysfunction with bleeding, and others. Macular edema threatening or involving the macular center is treated with focal macular photocoagulation. Small (50 μ in diameter), mild-intensity laser burns are targeted at areas of leakage in the macula (Murphy, *Amer. Family Physician* 51(4): 785-796 (1995)). If the macular edema persists, retreatment may be
30 necessary.

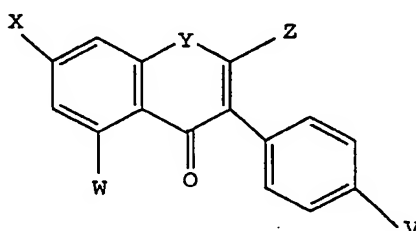
Patients with severe to very severe nonproliferative retinopathy and patients, who are at high risk for proliferative retinopathy or who already have early or advanced proliferative retinopathy, are treated with scatter or panretinal photocoagulation. Panretinal photocoagulation involves 1,500-2,000 laser burns, which are 500 μ in diameter, in the midperipheral and peripheral portion of the retina (Murphy (1995), *supra*).

The best documented biochemical mechanism for the development of microvascular complications of diabetes is the sorbitol pathway. In the sorbitol pathway, the enzyme aldose reductase catalyzes the conversion of glucose to sorbitol and galactose to galactitol. Aldose reductase has a low substrate affinity for glucose. Accordingly, when glucose concentrations are normal, the pathway is inactive. During hyperglycemia, the sorbitol pathway becomes active. Activation of the sorbitol pathway is important for retinal pericytes, for example, which do not require insulin for glucose penetration. Similarly, retinal capillary cells appear to contain substantial amounts of aldose reductase (Ferris, *Hospital Practice* ____: 79-89 (1993)).

Given the prevalence of diabetic retinopathy and ocular inflammation, there remains a need for an effective prophylactic and therapeutic treatment of diabetic retinopathy and ocular inflammation. Accordingly, it is a principal object of the present invention to provide a method of prophylactically and therapeutically treating diabetic retinopathy, including treatment at the pre-diabetic retinopathy stage, the nonproliferative diabetic retinopathy stage, and the proliferative diabetic retinopathy stage. It is another principal object of the present invention to provide a method of prophylactically and therapeutically treating ocular inflammation, e.g., macular edema, such as that associated with disease, bacterial or viral infection, or ocular surgery, such as cataract surgery, retinal surgery, refractive surgery, and corneal surgery, e.g., corneal transplantation, and the like, and corneal transplant rejection, among others. These and other objects of the present invention will become apparent from the detailed description provided herein.

BRIEF SUMMARY OF THE INVENTION

The present invention is directed to a method for the prophylactic and therapeutic treatment of diabetic retinopathy, including treatment at the pre-diabetic retinopathy stage, the nonproliferative diabetic retinopathy stage, and the proliferative diabetic retinopathy stage. The present invention is also directed to a method for the prophylactic and therapeutic treatment of ocular inflammation. The method involves the administration of an inhibitor of the protein tyrosine kinase pathway. Preferably, the inhibitor of the protein tyrosine kinase pathway is a compound of formula:



wherein V, W and X are selected from the group consisting of hydro, hydroxyl, alkoxy, halo, an ester, an ether, a carboxylic acid group, a pharmaceutically acceptable salt of a carboxylic acid group, and -SR, in which R is hydrogen or an alkyl group, and Y is selected from the group consisting of oxygen, sulfur, C(OH), and C=O, and Z is selected from the group consisting of hydro and C(O)OR₁, wherein R₁ is an alkyl. Preferably, the alkoxy is a C₁-C₆ alkoxy. Preferably, the halo is fluorine, chlorine or bromine. Preferably, the ester is a C₁-C₆ ester. Preferably, the ether is a C₁-C₆ ether. Preferred pharmaceutically acceptable salts of the carboxylic acid group include sodium and potassium salts. Preferably, the alkyl groups are C₁-C₆ alkyl groups. Desirably, the protein tyrosine kinase pathway inhibitor is genistein.

DETAILED DESCRIPTION OF THE INVENTION

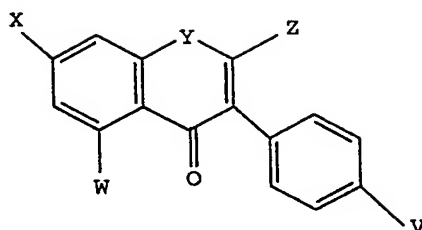
The present invention is predicated on the discovery that tyrosine phosphorylation is increased in the early stages of non-ischemic microvascular diabetic retinopathy and that an inhibitor of the protein tyrosine kinase pathway, specifically genistein, is effective in preventing diabetic retinopathy. Accordingly, the present invention provides a method for the prophylactic and therapeutic treatment of

diabetic retinopathy, including treatment at the pre-diabetic retinopathy stage, the nonproliferative diabetic retinopathy stage, and the proliferative diabetic retinopathy stage. In addition, the present invention provides a method for the prophylactic and therapeutic treatment of ocular inflammation. By "prophylactic" is meant the protection, in whole or in part, against diabetic retinopathy, in particular diabetic macular edema, or ocular inflammation. By "therapeutic" is meant the amelioration of diabetic retinopathy or ocular inflammation, itself, and the protection, in whole or in part, against further diabetic retinopathy or ocular inflammation. The present inventive method for the prophylactic and therapeutic treatment of ocular inflammation is particularly useful in the treatment of ocular inflammation, e.g., macular edema, due to disease, bacterial or viral infection, or ocular surgery, such as cataract surgery, retinal surgery, refractive surgery, and corneal surgery, e.g., corneal transplantation, and the like, and corneal transplant rejection, among others. In this regard, the present inventive method is also useful in the treatment of retinal edema associated with, for example, postcataract or laser capsulotomy (Irvine-Gass syndrome), uveitis, branch or central vein occlusion, topical epinephrine use, severe hypertension, radiation retinopathy, perifoveal telangiectasia and retinitis pigmentosa.

The method comprises the administration of an inhibitor of the protein tyrosine kinase pathway in an amount sufficient to treat ocular inflammation or the retina for retinopathy prophylactically or therapeutically. Any inhibitor of the protein tyrosine kinase pathway can be used in the method of the present invention as long as it is safe and efficacious. Herein, "PTK inhibitor" will be used to refer to such compounds and is intended to encompass all compounds that affect the protein tyrosine kinase pathway at any and all points in the pathway.

Preferably, the PTK inhibitor is genistein (5,7-dihydroxy-3-(4-hydroxyphenyl)-4H-1-benzopyran-4-one) or a pharmaceutically acceptable, protein tyrosine kinase pathway-inhibiting analogue or prodrug thereof or a pharmaceutically acceptable salt of any of the foregoing. Accordingly, the PTK inhibitor can be a compound of the following formula:

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wherein V, W and X are selected from the group consisting of hydro, hydroxyl, alkoxy, halo, an ester, an ether, a carboxylic acid group, a pharmaceutically acceptable salt of a carboxylic acid group, and -SR, in which R is hydrogen or an alkyl group, and Y is selected from the group consisting of oxygen, sulfur, C(OH), and C=O, and Z is selected from the group consisting of hydro and C(O)OR₁, wherein R₁ is an alkyl. Preferably, the alkoxy is a C₁-C₆ alkoxy. Preferably, the halo is fluorine, chlorine or bromine. Preferably, the ester is a C₁-C₆ ester. Preferably, the ether is a C₁-C₆ ether.

Preferred pharmaceutically acceptable salts of the carboxylic acid group include sodium and potassium salts. Preferably, the alkyl groups are C₁-C₆ alkyl groups. Desirably, the protein tyrosine kinase pathway inhibitor is genistein.

The prodrug can be any pharmaceutically acceptable prodrug of genistein, a protein tyrosine kinase pathway-inhibiting analogue of genistein, or a pharmaceutically acceptable salt of either of the foregoing. One of ordinary skill in the art will appreciate, however, that the prodrug used must be one that can be converted to an active PTK inhibitor in or around the retina. A preferred prodrug is a prodrug that increases the lipid solubility of genistein, a protein tyrosine kinase pathway-inhibiting analogue of genistein, or a pharmaceutically acceptable salt of either of the foregoing. A preferred prodrug is one in which one or more of V, W and X are independently derivatized with an ester, such as pivalic acid.

Compounds of the above formula are widely available commercially. For example, genistein is available from LC Laboratories (Woburn, MA). Those compounds that are not commercially available can be readily prepared using organic synthesis methods known in the art.

Whether or not a particular analogue, prodrug or pharmaceutically acceptable salt of a compound in accordance with the present invention can treat retinopathy

prophylactically or therapeutically can be determined by its effect in the rat model used in Example 1. Alternatively, analogues, prodrugs and pharmaceutically acceptable salts of inhibitors of the protein tyrosine kinase pathway can be tested by *in vitro* studies of endothelial cell proliferation and in other models of diabetic
5 retinopathy, such as Streptozotocin. Analogues, prodrugs and pharmaceutically acceptable salts of inhibitors of the protein tyrosine kinase pathway can be tested by clinical examination, such as by quantitation of cellular and humoral mediators of inflammation, or using the methods set forth in Example 2 for efficacy in the prophylactic and therapeutic treatment of ocular inflammation.

10 In addition, color Doppler imaging can be used to evaluate the action of a drug in ocular pathology (Valli et al., *Ophthalmologica* 209(13): 115-121 (1995)). Color Doppler imaging is a recent advance in ultrasonography, allowing simultaneous two-dimension imaging of structures and the evaluation of blood flow. Accordingly, retinopathy and ocular inflammation can be analyzed using such technology.

15 The PTK inhibitor can be bound to a suitable matrix, such as a polymeric matrix, if desired, for use in the present inventive method. Any of a wide range of polymers can be used in the context of the present invention provided that, if the polymer-bound compound is to be used *in vivo*, the polymer is biologically acceptable (see, e.g., U.S. Patent Nos. 5,384,333 and 5,164,188).

20 An advantage of genistein is that it is very safe and efficacious. For example, when genistein was orally administered to Zucker diabetic fatty rats, genistein was found to be nontoxic to the retina at dosages ranging from 75 mg/kg/day to 300 mg/kg/day over a period of six months as measured by electroretinography. In addition, oral administration of genistein was found to have no effect on food intake
25 and body weight for male and female rats. Also, no effect of orally administered genistein was found with respect to the weight of the ovaries and the uterus in female rats.

The PTK inhibitor, which is preferably genistein, a protein tyrosine kinase pathway-inhibiting analogue of genistein, a protein tyrosine kinase pathway-inhibiting
30 prodrug of genistein, or a pharmaceutically acceptable salt of any of the foregoing, can be administered in accordance with the present inventive method by any suitable

route. Suitable routes of administration include systemic, such as orally or by injection, topical, intraocular, periocular (e.g., subTenon's), subconjunctival, subretinal, suprachoroidal and retrobulbar. The manner in which the PTK inhibitor is administered is dependent, in part, upon whether the treatment of retinopathy or ocular inflammation is prophylactic or therapeutic. The manner in which the PTK inhibitor is administered for therapeutic treatment of retinopathy or ocular inflammation is dependent, in part, upon the cause of the retinopathy or ocular inflammation, respectively.

For example, given that diabetes is the leading cause of retinopathy, the PTK inhibitor can be administered prophylactically as soon as the pre-diabetic retinopathy state is detected. For the prophylactic treatment of retinopathy that can result from diabetes, the PTK inhibitor is preferably administered systemically, e.g., orally or by injection. For the therapeutic treatment of nonproliferative diabetic retinopathy, the PTK inhibitor can be administered systemically, e.g., orally or by injection, or intraocularly. Proliferative diabetic retinopathy can be therapeutically treated by the administration of the PTK inhibitor intraocularly, topically, subconjunctivally or periocularly (e.g., subTenon's), for example. The PTK inhibitor is preferably administered intraocularly, topically, subconjunctivally or periocularly (e.g., subTenon's) for the prophylactic or therapeutic treatment of retinopathy before, during or after surgical removal from an eye of scar tissue generated during neovascularization during the proliferative diabetic stage.

Similarly, since disease, bacterial or viral infection, ocular surgery, such as cataract surgery, retinal surgery, refractive surgery, and corneal surgery, e.g., corneal transplantation, and the like, and corneal transplant rejection, among others, are major causes of ocular inflammation, e.g., macular edema, the PTK inhibitor can be administered therapeutically as soon as disease or infection is detected or ocular surgery is completed. The PTK inhibitor can be administered prophylactically before disease or infection is detected or before, during or after ocular surgery. For the prophylactic treatment of ocular inflammation, the PTK inhibitor is preferably administered intraocularly or subretinally. In this regard, the use of an intraocular

device/implant, such as Vitrasert™, that is bioerodible and bioabsorbable, can be used to achieve sustained release of the PTK inhibitor.

The PTK inhibitor is preferably administered as soon as possible after it has been determined that an animal, such as a mammal, specifically a human, is at risk for
5 retinopathy or ocular inflammation (prophylactic treatment) or has begun to develop retinopathy or ocular inflammation (therapeutic treatment). Treatment will depend, in part, upon the particular PTK inhibitor used, the amount of the PTK inhibitor administered, the route of administration, and the cause and extent, if any, of retinopathy or ocular inflammation realized.

10 One skilled in the art will appreciate that suitable methods of administering a PTK inhibitor, which is useful in the present inventive method, are available. Although more than one route can be used to administer a particular PTK inhibitor, a particular route can provide a more immediate and more effective reaction than another route. Accordingly, the described routes of administration are merely
15 exemplary and are in no way limiting.

The dose administered to an animal, particularly a human, in accordance with the present invention should be sufficient to effect the desired response in the animal over a reasonable time frame. One skilled in the art will recognize that dosage will depend upon a variety of factors, including the strength of the particular PTK inhibitor
20 employed, the age, species, condition or disease state, and body weight of the animal, as well as the amount of the retina about to be affected or actually affected by retinopathy or the amount and location of ocular inflammation. The size of the dose also will be determined by the route, timing and frequency of administration as well as the existence, nature, and extent of any adverse side effects that might accompany
25 the administration of a particular PTK inhibitor and the desired physiological effect. It will be appreciated by one of ordinary skill in the art that various conditions or disease states, in particular, chronic conditions or disease states, may require prolonged treatment involving multiple administrations.

Suitable doses and dosage regimens can be determined by conventional range-
30 finding techniques known to those of ordinary skill in the art. Generally, treatment is initiated with smaller dosages, which are less than the optimum dose of the

compound. Thereafter, the dosage is increased by small increments until the optimum effect under the circumstances is reached. The present inventive method will typically involve the administration of from about 1 mg/kg/day to about 100 mg/kg/day, preferably from about 15 mg/kg/day to about 50 mg/kg/day, if administered
5 systemically. Intraocular administration typically will involve the administration of from about 0.1 mg total to about 5 mg total, preferably from about 0.5 mg total to about 1 mg total. A preferred concentration for topical administration is 100 μ M.

Compositions for use in the present inventive method preferably comprise a pharmaceutically acceptable carrier and an amount of a PTK inhibitor sufficient to
10 treat retinopathy or ocular inflammation prophylactically or therapeutically. The carrier can be any of those conventionally used and is limited only by chemico-physical considerations, such as solubility and lack of reactivity with the compound, and by the route of administration. It will be appreciated by one of ordinary skill in the art that, in addition to the following described pharmaceutical compositions, the
15 PTK inhibitor can be formulated as polymeric compositions, inclusion complexes, such as cyclodextrin inclusion complexes, liposomes, microspheres, microcapsules and the like (see, e.g., U.S. Patent Nos. 4,997,652, 5,185,152 and 5,718,922).

The PTK inhibitor can be formulated as a pharmaceutically acceptable acid addition salt. Examples of pharmaceutically acceptable acid addition salts for use in
20 the pharmaceutical composition include those derived from mineral acids, such as hydrochloric, hydrobromic, phosphoric, metaphosphoric, nitric and sulfuric acids, and organic acids, such as tartaric, acetic, citric, malic, lactic, fumaric, benzoic, glycolic, gluconic, succinic, and arylsulphonic, for example p-toluenesulphonic, acids.

The pharmaceutically acceptable excipients described herein, for example,
25 vehicles, adjuvants, carriers or diluents, are well-known to those who are skilled in the art and are readily available to the public. It is preferred that the pharmaceutically acceptable carrier be one which is chemically inert to the PTK inhibitor and one which has no detrimental side effects or toxicity under the conditions of use.

The choice of excipient will be determined in part by the particular PTK
30 inhibitor, as well as by the particular method used to administer the composition. Accordingly, there is a wide variety of suitable formulations of the pharmaceutical

composition of the present invention. The following formulations are merely exemplary and are in no way limiting.

Injectable formulations are among those that are preferred in accordance with the present inventive method. The requirements for effective pharmaceutically carriers for injectable compositions are well-known to those of ordinary skill in the art (see Pharmaceutics and Pharmacy Practice, J.B. Lippincott Co., Philadelphia, PA, Banker and Chalmers, eds., pages 238-250 (1982), and ASHP Handbook on Injectable Drugs, Toissel, 4th ed., pages 622-630 (1986)). It is preferred that such injectable compositions be administered intramuscularly, intravenously, or intraperitoneally.

Topical formulations are well-known to those of skill in the art. Such formulations are suitable in the context of the present invention for application to the skin. The use of patches, corneal shields (see, e.g., U.S. Patent No. 5,185,152), and ophthalmic solutions (see, e.g., U.S. Patent No. 5,710,182) and ointments, e.g., eye drops, is also within the skill in the art.

Formulations suitable for oral administration can consist of (a) liquid solutions, such as an effective amount of the compound dissolved in diluents, such as water, saline, or orange juice; (b) capsules, sachets, tablets, lozenges, and troches, each containing a predetermined amount of the active ingredient, as solids or granules; (c) powders; (d) suspensions in an appropriate liquid; and (e) suitable emulsions. Liquid formulations may include diluents, such as water and alcohols, for example, ethanol, benzyl alcohol, and the polyethylene alcohols, either with or without the addition of a pharmaceutically acceptable surfactant, suspending agent, or emulsifying agent. Capsule forms can be of the ordinary hard- or soft-shelled gelatin type containing, for example, surfactants, lubricants, and inert fillers, such as lactose, sucrose, calcium phosphate, and corn starch. Tablet forms can include one or more of lactose, sucrose, mannitol, corn starch, potato starch, alginic acid, microcrystalline cellulose, acacia, gelatin, guar gum, colloidal silicon dioxide, croscarmellose sodium, talc, magnesium stearate, calcium stearate, zinc stearate, stearic acid, and other excipients, colorants, diluents, buffering agents, disintegrating agents, moistening agents, preservatives, flavoring agents, and pharmacologically compatible excipients. Lozenge forms can comprise the active ingredient in a flavor, usually sucrose and

acacia or tragacanth, as well as pastilles comprising the active ingredient in an inert base, such as gelatin and glycerin, or sucrose and acacia, emulsions, gels, and the like containing, in addition to the active ingredient, such excipients as are known in the art.

5 Formulations suitable for parenteral administration include aqueous and non-aqueous, isotonic sterile injection solutions, which can contain anti-oxidants, buffers, bacteriostats, and solutes that render the formulation isotonic with the blood of the intended recipient, and aqueous and non-aqueous sterile suspensions that can include suspending agents, solubilizers, thickening agents, stabilizers, and preservatives. The
10 inhibitor can be administered in a physiologically acceptable diluent in a pharmaceutical carrier, such as a sterile liquid or mixture of liquids, including water, saline, aqueous dextrose and related sugar solutions, an alcohol, such as ethanol, isopropanol, or hexadecyl alcohol, glycols, such as propylene glycol or polyethylene glycol, dimethylsulfoxide, glycerol ketals, such as 2,2-dimethyl-1,3-dioxolane-4-
15 methanol, ethers, such as poly(ethyleneglycol) 400, an oil, a fatty acid, a fatty acid ester or glyceride, or an acetylated fatty acid glyceride, with or without the addition of a pharmaceutically acceptable surfactant, such as a soap or a detergent, suspending agent, such as pectin, carbomers, methylcellulose, hydroxypropylmethylcellulose, or carboxymethylcellulose, or emulsifying agents and other pharmaceutical adjuvants.
20 Oils, which can be used in parenteral formulations include petroleum, animal, vegetable, or synthetic oils. Specific examples of oils include peanut, soybean, sesame, cottonseed, corn, olive, petrolatum, and mineral.

 Suitable fatty acids for use in parenteral formulations include oleic acid, stearic acid, and isostearic acid. Ethyl oleate and isopropyl myristate are examples of
25 suitable fatty acid esters.

 Suitable soaps for use in parenteral formulations include fatty alkali metals, ammonium, and triethanolamine salts, and suitable detergents include (a) cationic detergents such as, for example, dimethyl dialkyl ammonium halides, and alkyl pyridinium halides, (b) anionic detergents such as, for example, alkyl, aryl, and olefin
30 sulfonates, alkyl, olefin, ether, and monoglyceride sulfates, and sulfosuccinates, (c) nonionic detergents such as, for example, fatty amine oxides, fatty acid

alkanolamides, and polyoxyethylenepolypropylene copolymers, (d) amphoteric detergents such as, for example, alkyl-p-aminopropionates, and 2-alkyl-imidazoline quaternary ammonium salts, and (e) mixtures thereof.

5 The parenteral formulations will typically contain from about 0.5 to about 25% by weight of the active ingredient in solution. Preservatives and buffers may be used. In order to minimize or eliminate irritation at the site of injection, such compositions may contain one or more nonionic surfactants having a hydrophile-lipophile balance (HLB) of from about 12 to about 17.

10 The quantity of surfactant in such formulations will typically range from about 5 to about 15% by weight. Suitable surfactants include polyethylene sorbitan fatty acid esters, such as sorbitan monooleate and the high molecular weight adducts of ethylene oxide with a hydrophobic base, formed by the condensation of propylene oxide with propylene glycol. The parenteral formulations can be presented in unit-dose or multi-dose sealed containers, such as ampules and vials, and can be stored in a
15 freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid excipient, for example, water, for injections, immediately prior to use. Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules, and tablets of the kind previously described.

Such compositions can be formulated as intraocular formulations, sustained-
20 release formulations or devices (see, e.g., U.S. Patent No. 5,378,475). For example, gelatin, chondroitin sulfate, a polyphosphoester, such as bis-2-hydroxyethyl-terephthalate (BHET), or a polylactic-glycolic acid (in various proportions) can be used to formulate sustained-release formulations. Implants (see, e.g., U.S. Patent Nos. 5,443,505, 4,853,224 and 4,997,652), devices (see, e.g., U.S. Patent Nos. 5,554,187,
25 4,863,457, 5,098,443 and 5,725,493), such as an implantable device, e.g., a mechanical reservoir, an intraocular device or an extraocular device with an intraocular conduit (e.g., 100 μ - 1 mm in diameter), or an implant or a device comprised of a polymeric composition as described above, can be used.

The present inventive method also can involve the co-administration of other
30 pharmaceutically active compounds. By "co-administration" is meant administration before, concurrently with, e.g., in combination with the PTK inhibitor in the same

formulation or in separate formulations, or after administration of a PTK inhibitor as described above. For example, corticosteroids, e.g., prednisone, methylprednisolone, dexamethasone, or triamcinalone acetonide, or noncorticosteroid anti-inflammatory compounds, such as ibuprofen or flubiprofen, can be co-administered. Similarly, 5 vitamins and minerals, e.g., zinc, anti-oxidants, e.g., carotenoids (such as a xanthophyll carotenoid like zeaxanthin or lutein), and micronutrients can be co-administered. In addition, other types of inhibitors of the protein tyrosine kinase pathway, which include natural protein tyrosine kinase inhibitors like quercetin, lavendustin A, erbstatin and herbimycin A, and synthetic protein tyrosine kinase 10 inhibitors like tyrphostins (e.g., AG490, AG17, AG213 (RG50864), AG18, AG82, AG494, AG825, AG879, AG1112, AG1296, AG1478, AG126, RG13022, RG14620 and AG555), dihydroxy- and dimethoxybenzylidene malononitrile, analogs of lavendustin A (e.g., AG814 and AG957), quinazolines (e.g., AG1478), 4,5-dianilinophthalimides, and thiazolidinediones, can be co-administered with genistein 15 or an analogue, prodrug or pharmaceutically acceptable salt thereof (see Levitzki et al., *Science* 267: 1782-1788 (1995); and Cunningham et al., *Anti-Cancer Drug Design* 7: 365-384 (1992)). In this regard, potentially useful derivatives of genistein include those set forth in Mazurek et al., U.S. Patent No. 5,637,703. Neutralizing proteins to growth factors, such as a monoclonal antibody that is specific for a given growth 20 factor, e.g., VEGF (for an example, see Aiello et al., *PNAS USA* 92: 10457-10461 (1995)), or phosphotyrosine (Dhar et al., *Mol. Pharmacol.* 37: 519-525 (1990)), can be co-administered. Other various compounds that can be co-administered include inhibitors of protein kinase C (see, e.g., U.S. Patent Nos. 5,719,175 and 5,710,145), cytokine modulators, an endothelial cell-specific inhibitor of proliferation, e.g., 25 thrombospondins, an endothelial cell-specific inhibitory growth factor, e.g., $\text{TNF}\alpha$, an anti-proliferative peptide, e.g., SPARC and prolifeferin-like peptides, a glutamate receptor antagonist, aminoguanidine, an angiotensin-converting enzyme inhibitor, e.g., angiotensin II, calcium channel blockers, ψ -tectorigenin, ST638, somatostatin analogues, e.g., SMS 201-995, monosialoganglioside GM1, ticlopidine, neurotrophic 30 growth factors, methyl-2,5-dihydroxycinnamate, an angiogenesis inhibitor, e.g., recombinant EPO, a sulphonylurea oral hypoglycemic agent, e.g., glizalazide (non-

insulin-dependent diabetes), ST638 (Asahi et al., *FEBS Letter* 309: 10-14 (1992)), thalidomide, nicardipine hydrochloride, aspirin, piceatannol, staurosporine, adriamycin, epiderstatin, (+)-aeropylsinin-1, phenazocine, halomethyl ketones, anti-lipidemic agents, e.g., etofibrate, chlorpromazine and sphingosines, aldose reductase inhibitors, such as tolrestat, SPR-210, sorbinil or oxygen, and retinoic acid and analogues thereof (Burke et al., *Drugs of the Future* 17(2): 119-131 (1992); and Tomlinson et al., *Pharmac. Ther.* 54: 151-194 (1992)). Selenoindoles (2-thioindoles) and related disulfide selenides, such as those described in Dobrusin et al., U.S. Patent No. 5,464,961, are useful protein tyrosine kinase inhibitors. Those patients that exhibit systemic fluid retention, such as that due to cardiovascular or renal disease and severe systemic hypertension, can be additionally treated with diuresis, dialysis, cardiac drugs and antihypertensive agents.

EXAMPLES

The following examples further illustrate the present invention but, of course, should not be construed as in any way limiting its scope.

EXAMPLE 1

This example demonstrates that tyrosine phosphorylation is increased in the early stages of non-ischemic microvascular diabetic retinopathy and that genistein inhibits such tyrosine phosphorylation.

Six pair of litter mates of Zucker rats, i.e., six male retired breeder Zucker diabetic fatty (ZDF, fa/fa) rats and six male retired breeder lean Zucker (fa/+) rats, were obtained (Genetic Models, Inc., Indianapolis, IN). The Zucker diabetic fatty rat develops a microvascular retinopathy involving basement membrane thickening of retinal capillaries, pericyte loss, capillary hypercellularity, and increased endothelial intercellular junctions (Danis et al., *Invest. Ophthalmol. Vis. Sci.* 34: 2367-2371 (1993); and Dosso et al., *Diabetologia* 33: 137-144 (1990)). The microvascular changes known to occur in the Zucker diabetic fatty rats are associated with the elevated levels of VEGF and platelet-derived growth factor (PDGF) and with activation of PTK pathways, especially pathways that include phosphatidylinositol-3

kinase (PI3-K) and mitogen-activated protein kinase (MAPK). Each pair of litter mates was between six and seven months of age. Rats were fed with a diet of Purina 5008 rat chow. The animals were treated in accordance with The Association for Research in Vision and Ophthalmology (ARVO) Statement for the Use of Animals in
5 Ophthalmic and Vision Research. The average blood glucose value was more than 500 mg/dl for the Zucker diabetic fatty rats and 120 mg/dl for the control lean rats as measured by glucose oxidase assay at the time of death (Danis et al. (1993), *supra*).

After an intraperitoneal injection of ketamine (40 mg/kg) and xylazine (4 mg/kg), genistein (LC Laboratories, Woburn, MA), which was dissolved in DMSO at
10 the concentration of 10 mg/ml, was injected intraperitoneally into the Zucker diabetic fatty rat and lean rat (non-treated group) at a dose of 50 mg/kg body weight. After two hours, the injection was repeated.

Twenty four hours later, the animals were sacrificed by an injection of intracardiac pentobarbital. Then, 1 mM sodium orthovanadate was injected into the
15 rat eye to inhibit phosphatase activity, the eye was enucleated and the anterior segment was cut away. The vitreous was removed with forceps and the retina was peeled away from the retinal pigmented epithelium/choroid and cut from the optic nerve. The retinal tissue was placed into a tube with 125 μ l of ice-cold lysis buffer (150 mM NaCl, 1 % Triton X-100, 0.5 % sodium deoxycholate, 0.1 % SDS, 50 mM
20 Tris, 100 μ g/ml phenylmethylsulfonyl fluoride (PMSF), 0.3 μ g/ml EDTA, 0.7 μ g/ml pepstatin A, 0.5 μ g/ml leupeptin, 1 mM sodium orthovanadate, and 50 μ M sodium fluoride. The retinal tissue was homogenized with a glass homogenizer and then equal amounts of 2X SDS sample buffer (160 mM Tris-HCl, pH 6.8, 4 % SDS, 30 % glycerol, 5 B β -mercaptoethanol, 10 mM dithiothreitol, 0.01 % bromophenol blue)
25 were added. The retinal homogenate was boiled at 95°C for 5 min and centrifuged at 13,000 rpm for 10 min. The supernatants were collected and stored at -80°C. These total retinal samples were used for gel electrophoresis and Western immunoblot analysis.

Protein concentrations were measured by the Pierce BCA method. Total
30 protein profiles revealed that the Zucker diabetic fatty rat retina had a more strongly stained band at 67 kDa as compared with the lean controls. Treatment with genistein

reduced the amount of staining of the 67 kDa band in the Zucker diabetic fatty rat. Other bands in the total protein profiles appeared to be unchanged irrespective of the treatment with genistein.

Anti-phosphotyrosine antibody (PY 20; 1:500 dilution; Transduction
5 Laboratories, Lexington, KY), anti-PCNA antibody (1:200 dilution), anti-mitogen-
activated protein kinase antibody (1:100 dilution; Santa Cruz Biotechnology, Inc.,
Santa Cruz, CA), anti-phosphatidylinositol 3-kinase antibody (1:500 dilution; Upstate
Biotechnology, Inc., Lake Placid, NY) were used as primary antibodies in the Western
immunoblot analysis. Horseradish peroxidase (HRP) conjugated goat anti-mouse IgG
10 antibody or anti-rabbit IgG antibody (Bio-Rad Laboratories, Inc., Hercules, CA) was
used as the secondary antibody. Avidin-HRP (Bio-Rad) was used to detect
biotinylated molecular markers.

The retinal tissue samples were added to immunoprecipitation buffer (1 %
Triton X-100, 150 mM NaCl, 10 mM Tris, pH 7.4, 1 mM EGTA, 0.2 mM sodium
15 vanadate, 0.2 mM PMSF, 0.5 % NP-40) and homogenized. The supernatant was
obtained after centrifugation at 8,000 rpm for 10 min at 4°C. Protein concentrations
were measured by the Pierce BCA method. Then 20 µg of agarose-conjugated
phosphotyrosine antibody (PY 20; Transduction Laboratories) were added to each
milligram of retinal homogenate. Retinal homogenates were incubated with the
20 agarose-conjugated phosphotyrosine antibody at 4°C overnight with shaking. The
pellets were collected by centrifugation (10 min at 8,000 rpm) and rinsed with the
lysis buffer three times. The pellets were resuspended in 150 µl of 1X SDS sample
buffer per milligram of retinal homogenate and boiled at 95°C for 5 min. These
immunoprecipitated samples were used for gel electrophoresis.

25 Equal protein amounts of retinal samples were electrophoresed at a constant
200 V for 35 min on 4-20 % gradient minigels (Bio-Rad), using a Bio-Rad Protean II
apparatus. Broad range, biotinylated (6.5 - 200 kDa) and prestained (18.5 - 106 kDa)
molecular weight protein markers (Bio-Rad) were run simultaneously with the
samples. After electrophoresis, gels were processed either for total protein staining
30 with Coomassie brilliant blue dye or for Western immunoblot analysis.

After gel electrophoresis, the gels were fixed in 45% methanol and 10 % acetic acid aqueous solution for 30 min. Then, they were soaked in saturated picric acid solution briefly and stained with 0.25 % aqueous Coomassie brilliant blue (R-250) for a minimum of 2 hr. The gels were destained in 10 % acetic acid solution. The
5 procedure was repeated three times.

For Western immunoblot analysis, the gels were electroblotted onto nitrocellulose membranes (Coster Scientific Corp. Cambridge, MA) using a trans-blot SD apparatus (Bio-Rad). The membranes for detection of tyrosine-phosphorylated proteins were incubated with 3 % bovine serum albumin in Tris-buffered saline (TBS;
10 20 mM Tris and 150 mM NaCl, pH 7.5) for 1 hr at room temperature. The membranes for detection of other proteins were incubated with 2 % of nonfat dried milk (Bio-Rad) in TBS for 1 hr at room temperature. Then, each membrane was incubated with the primary antibody solution overnight at 4°C. Then, the membranes were rinsed with TBS and incubated with HRP-conjugated rat anti-mouse IgG (Jackson
15 Immunoresearch Laboratories, Inc., West Grove, PA) or anti-rabbit IgG antibody (Bio-Rad; 1:1,000 dilution) and avidin-HRP (Bio-Rad; 1:10,000 dilution) for 2 hr at room temperature. Following a final rinse series with TBS containing 0.2 % Tween-20 and TBS alone, the membranes were reacted with enhanced chemiluminescence (ECL) immunodetection reagents (Amersham Life Science Inc., Arlington Heights,
20 IL) and exposed to X-ray film. Each Western immunoblot analysis was repeated at least three times.

Overall, the tyrosine phosphorylated proteins in the diabetic retina were increased in comparison to the lean controls at 50, 66, 97 and 116 kDa. Genistein treated diabetic rats showed decreased staining at each molecular weight as compared
25 with the non-treated group.

Western immunoblot analysis of immunoprecipitated tyrosine phosphorylated proteins revealed the increased tyrosine phosphorylation of PI3-K at 85 kDa in the diabetic rats as compared to the lean controls. The tyrosine phosphorylated PI3-K stained band was reduced after treatment with genistein, especially in the diabetic rat.

Tyrosine phosphorylated MAPK at 44 kDa was increased in the diabetic rats as compared with the lean controls. Similar to PI3-K, the level of MAPK in the treated group was also decreased as compared with the non-treated animals group.

Two pairs of rats were used for immunohistochemical analysis of PCNA in the retina. Under deep anesthesia with ketamine and xylazine, the four rats were perfused transcardially with 0.1 M phosphate-buffered saline (PBS; pH 7.4) containing 1 mM orthovanadate, followed by perfusion of 2 % paraformaldehyde in 0.1 M PBS for fixation. The eyes were then removed and immersed in the same fixative for 30 min. The anterior segments were removed and the posterior segments were transferred into a graded sucrose solution from 5 % to 15 % for 2 hrs and incubated in 20 % sucrose solution overnight at 4°C. After cryoprotection, the eyes were molded in O.C.T. compound (Miles, Inc., Elkhart, IN) and frozen in isopentane with dry ice. The posterior segments were cut into 8 µm sections.

The sections were treated with 3 % peroxide to block intrinsic peroxidase, and incubated with 2 % normal horse serum for 20 min. After rinsing in 0.1 M PBS three times, the sections were incubated overnight at 4°C with anti-PCNA antibody (1:1,000 dilution). After the incubation, the sections were rinsed in 0.1 M PBS, and incubated with biotinylated anti-mouse IgG antibody (Vector Laboratories, Inc., Burlingame, CA; 1:500 dilution) for 30 min at room temperature, and then incubated with peroxide-labeled streptavidin (Kirkegaard & Perry Laboratories, Inc., Gaithersburg, MD; 1:500 dilution). Slides were treated with 3-amino-9-ethylcarbazole (AEC) solution (Sigma Chemical Co., St. Louis, MO) to enable visualization. Each set of slides was examined at the same time and three different sets were studied. After mounting with glycerin jelly (7 % gelatin, 54 % glycerine, 0.7 % phenol), photomicrographs were taken with Nomarski optics (Carl Zeiss, Thornwood, NY).

In the Zucker diabetic fatty rat retina, the level of PCNA at 36 kDa was increased as compared with the lean rats. Genistein reduced this increase as compared to the non-treated group, although not down to the lean control level.

An intense positive immunoreaction in the nuclei of the cells in the nerve fiber and the inner and outer nuclear layers was observed in the Zucker diabetic fatty rats

when compared with the lean controls. However, in the genistein treated diabetic rats, fewer immunopositive cells were detected. Incubation of the diabetic rat retina with non-immune mouse IgG, instead of anti-PCNA antibody, showed no immunoreaction.

The above results show that genistein reduced increased levels of phosphotyrosine overall and reduced increased levels of phosphorylated PI3-K, MAPK and PCNA (which is indicative of the cells being primed for replication) in the diabetic rat retina. These results suggest that genistein can inhibit the activation of PTK pathways in the retina of diabetic animals with pre-visual changes such that diabetic retinopathy can be prevented.

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EXAMPLE 2

This example demonstrates the ability of an inhibitor of the protein tyrosine kinase pathway to prophylactically and therapeutically treat ocular inflammation.

Herbimycin A (HA) is a benzenoid ansamycin antibiotic produced by the *Streptomyces hygroscopicus* strain. It binds tightly to the catalytic portion of the tyrosine kinase receptor, and is a potent and specific inhibitor of protein tyrosine phosphorylation, preventing phosphorylation of the protein messengers and subsequent activation of the cell and inhibiting cell proliferation.

The principles of laboratory animal care (NIH publication No. 86-23, revised 1985) were followed. For all studies, pigmented rabbits, weighing 2.5-3 kg were used. The animals were anesthetized with a cocktail of ketamine (25 mg/kg) and xylazine (5 mg/kg), and the pupils were dilated with topical 1% tropicamide and 0.5% phenylephrine.

Vitreous liquefaction was caused by injection of SF₆ gas. After an anterior chamber paracentesis (0.1 ml), pure SF₆ gas (0.3 ml) was injected. The 30-gauge needle was inserted inferiorly in order to maximize the vitreous liquefaction. We did not place cryopexy or perform gas-fluid exchange, thus circumventing associated complications.

Rabbit dermal fibroblasts were obtained from rabbit rump biopsy material prepared under sterile conditions: Primary cultures were incubated in Dulbecco's modified Eagle's medium (DMEM) with 30% calf serum, antibiotics (penicillin G

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sodium, streptomycin sulfate) and antimycotics (amphotericin B) under a humid atmosphere of 5% carbon dioxide in air for 72 hours. The medium was then changed to DMEM with 10% calf serum, antibiotics and antimycotics as above. The cultures were passaged weekly. Cells were harvested by incubating them with 3.5 ml of 0.04% trypsin for 4 minutes and collecting the cells in stop media. The dispersed cells were centrifuged at 1,000 rpm for 5 minutes and resuspended in 1 ml of phosphate-buffered saline. After cell counts of a 0.05 ml aliquot, the suspension was diluted to achieve a final concentration of 2.5×10^4 cells in a 0.1 ml of phosphate-buffered saline.

Fourteen days after the injection of SF6 gas, a 30-gauge needle was inserted 3 mm posterior to the corneoscleral limbus in the superonasal quadrant under microscopic control. With the bevel of the needle directed upward, 2.5×10^4 homologous rabbit dermal fibroblasts suspended in 0.1 ml sterile phosphate-buffered saline were injected just in front of the optic nerve head. This was performed in both eyes of all rabbits. The rabbits were immediately placed on their backs for 1 hour to allow the cells to settle over the vitreous surface.

In group 1, one eye in each rabbit ($n=18$) was injected with 23 μ g HA in 0.1 ml phosphate-buffered saline 1 hour (prophylactic) after fibroblast injection in order to achieve a final intraocular concentration of approximately 20 μ g HA. The fellow eye in each rabbit was injected with 0.1 ml phosphate-buffered saline as the control eye. In group 2, one eye in each rabbit ($n=10$) was injected with 23 μ g HA in 0.1 ml phosphate-buffered saline 2 days (therapeutic) after fibroblast injection, while the fellow eye was injected with 0.1 ml phosphate-buffered saline and served as control. In group 3, one eye in each rabbit ($n=9$) was injected with 11.5 μ g of HA (10 μ g in the eye) at 1 hour and again 2 days after fibroblast injection, while 0.1 ml phosphate-buffered saline was injected into the fellow eye.

Each eye was examined with indirect ophthalmoscopy and had fundus photographs taken 3 days after the fibroblast injection. Inflammation, as judged by vitreous cavity haze, was graded as mild or severe by means of fundus photographs. Severe inflammation caused sufficient haze in the photograph to prevent visualization of the myelinated "brush boarder" of the medullary wing near the optic nerve head.

Three days after fibroblast injection, the severity of inflammation (as judged by vitreous haze) was significantly different between control and treated eyes. In group 1, severe inflammation was present in 47.1% of the control eyes versus 0% of the treated eyes ($p < 0.01$). In group 3, severe inflammation was present in 77.8% of control eyes versus 22.2% of treated eyes ($p < 0.05$). In group 2, severe inflammation was present in 50% of control eyes versus 10% of treated eyes ($p > 0.05$).

In summary, vitreous haze, an indicator of inflammation, was decreased in the eyes that were treated with HA. This example demonstrates the ability of an inhibitor of the protein tyrosine kinase pathway to treat ocular inflammation prophylactically and therapeutically.

All of the references cited herein, including patents, patent applications, literature publications, and the like, are hereby incorporated in their entireties by reference.

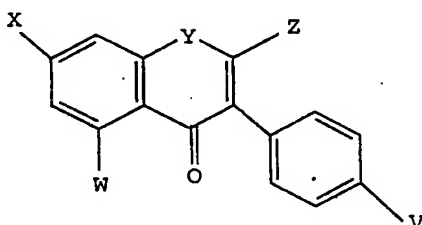
While this invention has been described with an emphasis upon preferred embodiments, it will be obvious to those of ordinary skill in the art that variations of the preferred compounds and methods may be used and that it is intended that the invention may be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the invention as defined by the following claims.

What is claimed is:

1. A method of prophylactically or therapeutically treating an animal for diabetic retinopathy, which method comprises administering to said animal an inhibitor of the protein tyrosine kinase pathway in an amount sufficient to treat said
5 animal for diabetic retinopathy prophylactically or therapeutically.

2. A method of prophylactically or therapeutically treating an animal for ocular inflammation, which method comprises administering to said animal an inhibitor of the protein tyrosine kinase pathway in an amount sufficient to treat said
10 animal for ocular inflammation prophylactically or therapeutically.

3. The method of claim 1 or 2, wherein said inhibitor of the protein tyrosine kinase pathway is a compound of formula:



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wherein V, W and X are selected from the group consisting of hydro, alkoxy, hydroxyl, halo, an ester, an ether, a carboxylic acid group, a pharmaceutically acceptable salt of a carboxylic acid group, and -SR, in which R is hydrogen or an
20 alkyl group, and Y is selected from the group consisting of oxygen, sulfur, C(OH), and C=O, and Z is selected from the group consisting of hydro and C(O)OR₁, wherein R₁ is an alkyl, or a protein tyrosine kinase-inhibiting prodrug or pharmaceutically acceptable salt thereof.

25 4. The method of claim 3, wherein the halo group is selected from the group consisting of fluorine, chlorine and bromine.

5. The method of claim 3, wherein the ester is a C₁-C₆ ester.
6. The method of claim 3, wherein the ether is a C₁-C₆ ether.
- 5 7. The method of claim 3, wherein said pharmaceutically acceptable salt of a carboxylic acid group is a sodium salt or a potassium salt.
8. The method of claim 3, wherein the alkyl groups are C₁-C₆ alkyl groups and the alkoxy group is a C₁-C₆ alkoxy group.
- 10 9. The method of claim 3, wherein said inhibitor of the protein tyrosine kinase pathway is genistein.
10. The method of any of claims 1 and 3-9, wherein genistein is administered
- 15 systemically.
11. The method of claim 10, wherein genistein is administered in an amount from about 1 mg/kg/day to about 100 mg/kg/day.
12. The method of claim 11, wherein genistein is administered in an amount
- 20 from about 15 mg/kg/day to about 50 mg/kg/day.
13. The method of claim 10, wherein genistein is administered orally or by injection.
- 25 14. The method of claim 10, wherein genistein is administered at the pre-diabetic retinopathy stage.
15. The method of claim 10, wherein genistein is administered at the
- 30 nonproliferative diabetic retinopathy stage.

16. The method of claim 15, wherein the administration of genistein prevents diabetic macular edema.

17. The method of claim 15, wherein the administration of genistein treats
5 diabetic macular edema.

18. The method of claim 10, wherein genistein is administered at the proliferative diabetic stage.

19. The method of claim 18, wherein genistein is administered before, during
10 or after surgical removal from an eye of scar tissue generated during neovascularization during the proliferative diabetic stage.

20. The method of any of claims 2-9, wherein said ocular inflammation is that
15 which results from a disease.

21. The method of any of claims 2-9, wherein said ocular inflammation is that which results from a bacterial or viral infection.

22. The method of any of claims 2-9, wherein said ocular inflammation is that
20 which results from ocular surgery.

23. The method of claim 22, wherein said ocular surgery is cataract surgery, retinal surgery, refractive surgery, or corneal surgery.

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24. The method of claim 23, wherein said corneal surgery is corneal transplantation.

25. The method of claim 24, wherein said corneal transplantation results in
30 corneal transplant rejection.

26. The method of any of claims 2-9, wherein said ocular inflammation is nondiabetic macular edema.

27. The method of any of claims 1-26, wherein genistein is administered
5 topically, subconjunctivally, retrobulbarly, periocularly, subretinally, suprachoroidally or intraocularly.

28. The method of any of claims 3-9 and 20-27, wherein genistein is administered intraocularly in an amount from about 0.1 mg total to about 5 mg total.

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29. The method of claim 28, wherein genistein is administered intraocularly in an amount from about 0.5 mg total to about 1 mg total.

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